

A multi-omic approach to uncover enhancer-gene interactions in the human brain

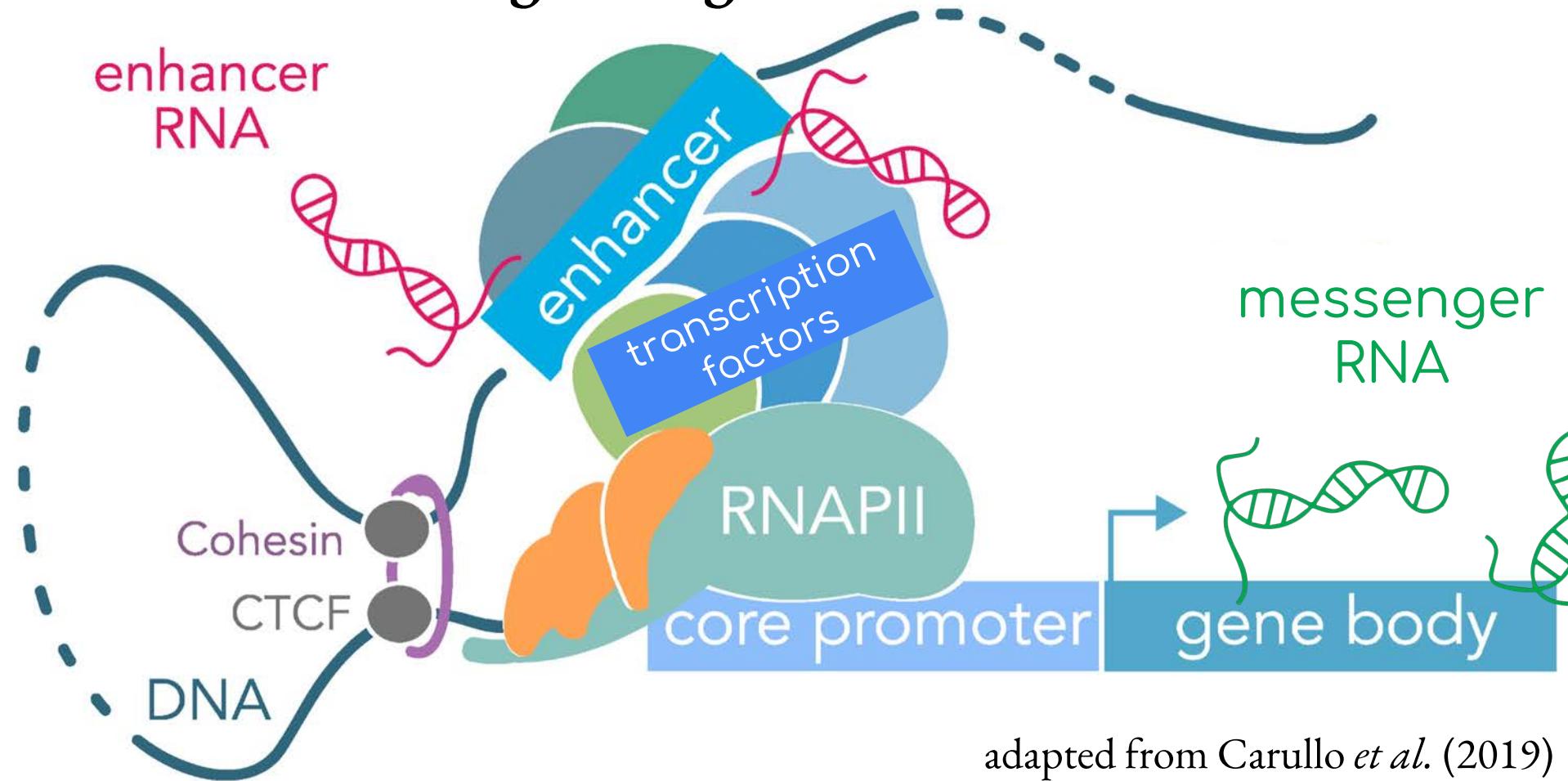
MIT Primes Conference

10.12.24



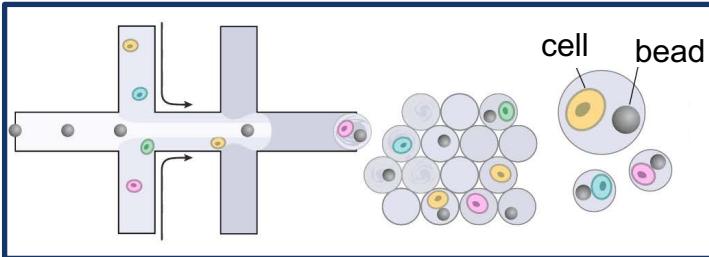
Sophia Yan
(Steve McCarroll Lab)

Enhancer-driven gene regulation

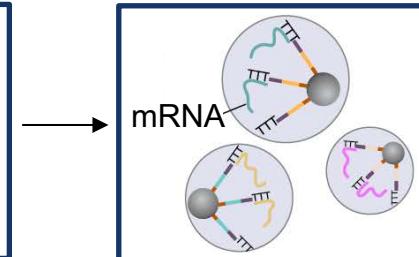


Single nucleus RNA sequencing (snRNA-seq)

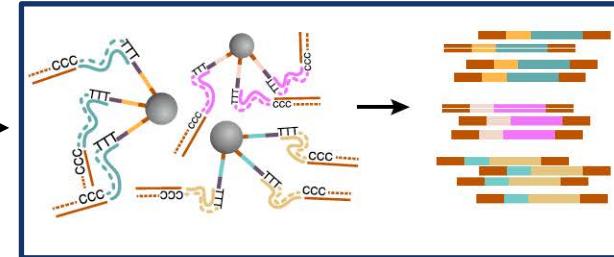
1. Droplet Formation



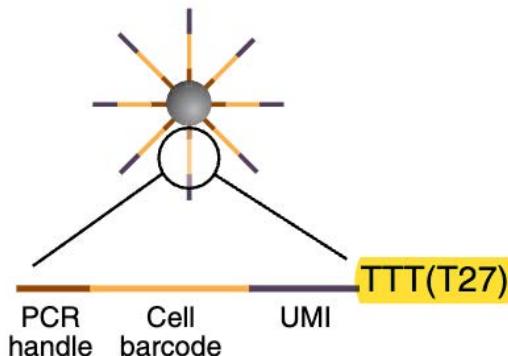
2. Cell Lysis (in seconds)



3. Polymerase Chain Reaction



Barcoded primer bead



Cell Barcode

TTGCCGTGGTGTGGCGGGGA.....CGGTCTTA

TTGCCGTGGTGTATGGAGG.....AAAATGGC

TTGCCGTGGTGTATGGAGG.....AAAATGGC

UMI

CGTTAGATGGCA.....CTCATAGT

CGTTAGATGGCAACGTTATA.....ACCGTAC

mRNA read

GTTAACGTACCTGTGCTTG.....CCAGCACC

GTTAACGTACCTGTGCTTG.....CCAGCACC

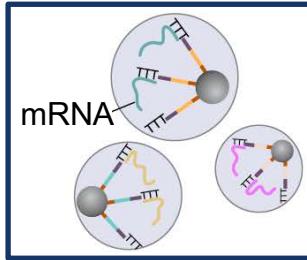
GTTAACGTACCTGTGCTTG.....CCAGCACC

GTTAACGTACCCTAGCTGT.....TTCCGGTC

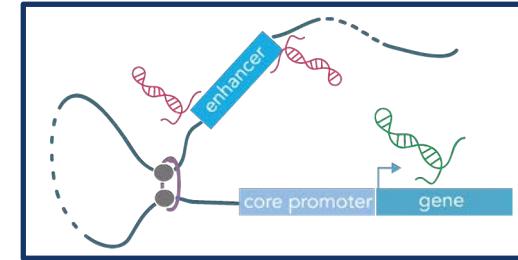
Project overview

Objectives

- a) Detect eRNA from snRNA-seq data



- b) Link putative active enhancers back to their target genes



Motivation

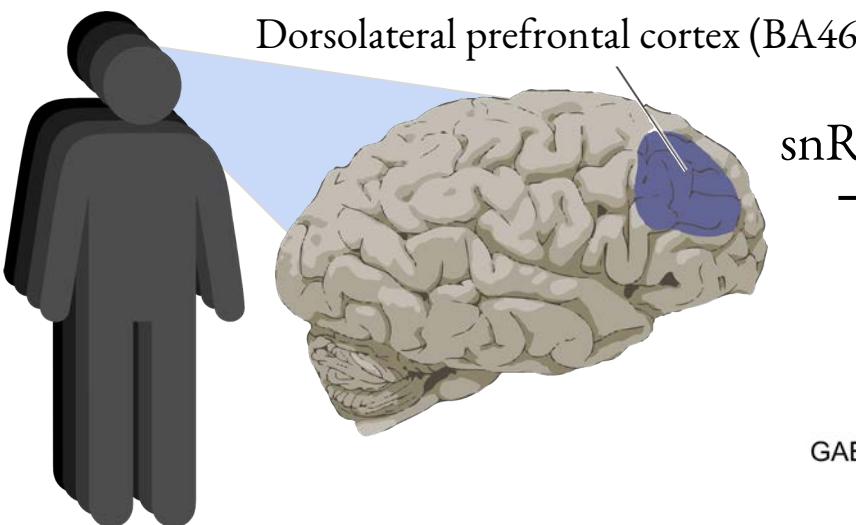
- a) Ability to explore active enhancers at the cell-type specific level

Challenges

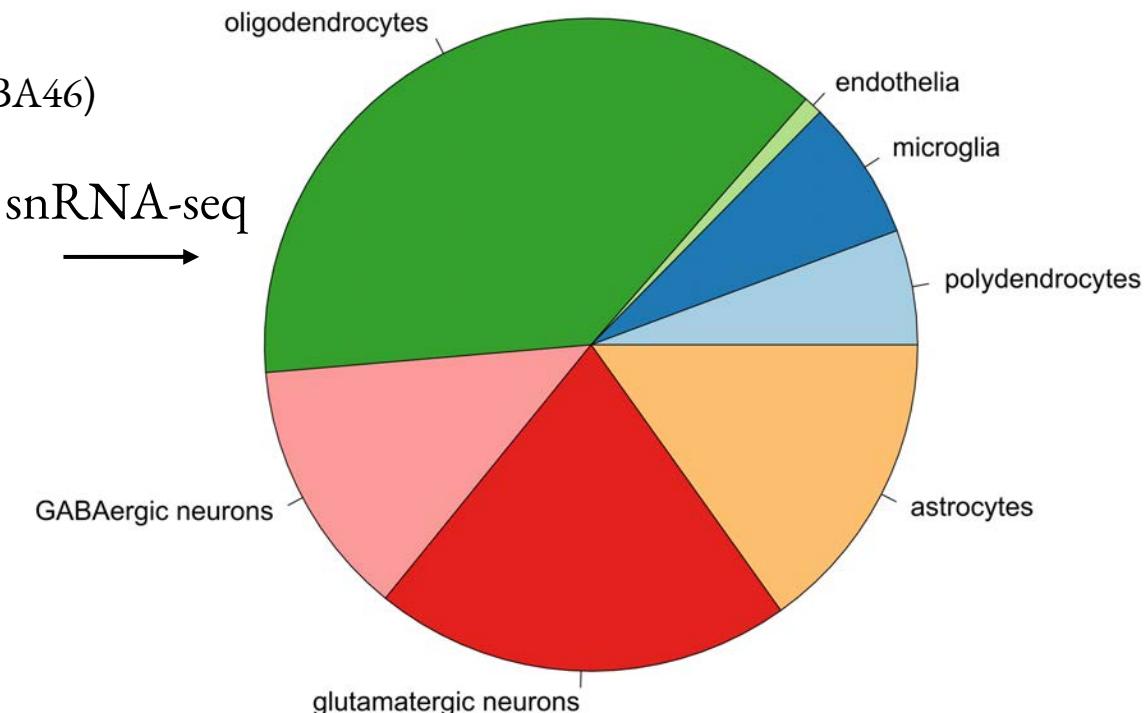
- a) eRNA is unstable
- b) snRNA-seq is not optimized to target eRNA

Our snRNA-seq dataset

20 donors



Cell-type composition of
79k total nuclei

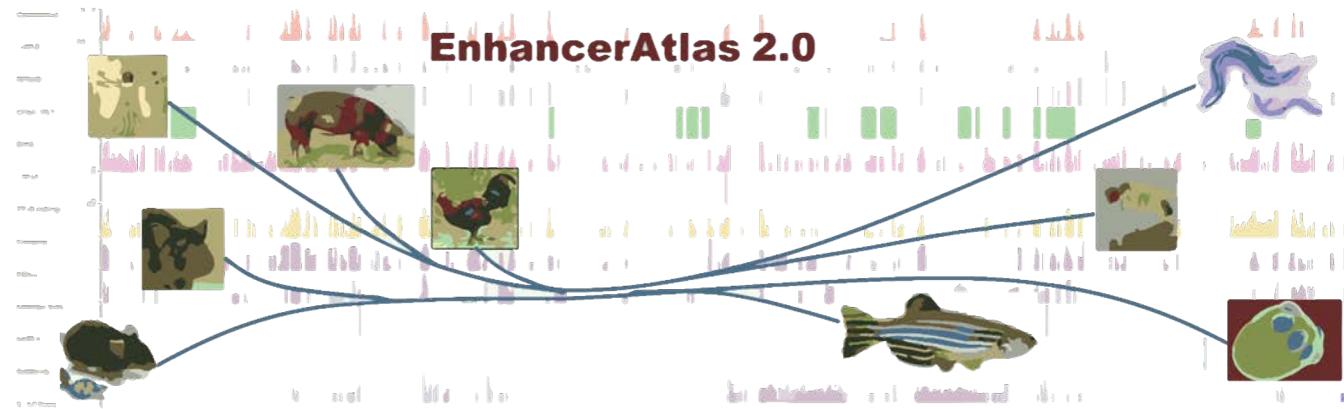


Identifying 29k putative brain enhancers from reference databases

62k brain enhancers



129k brain enhancers

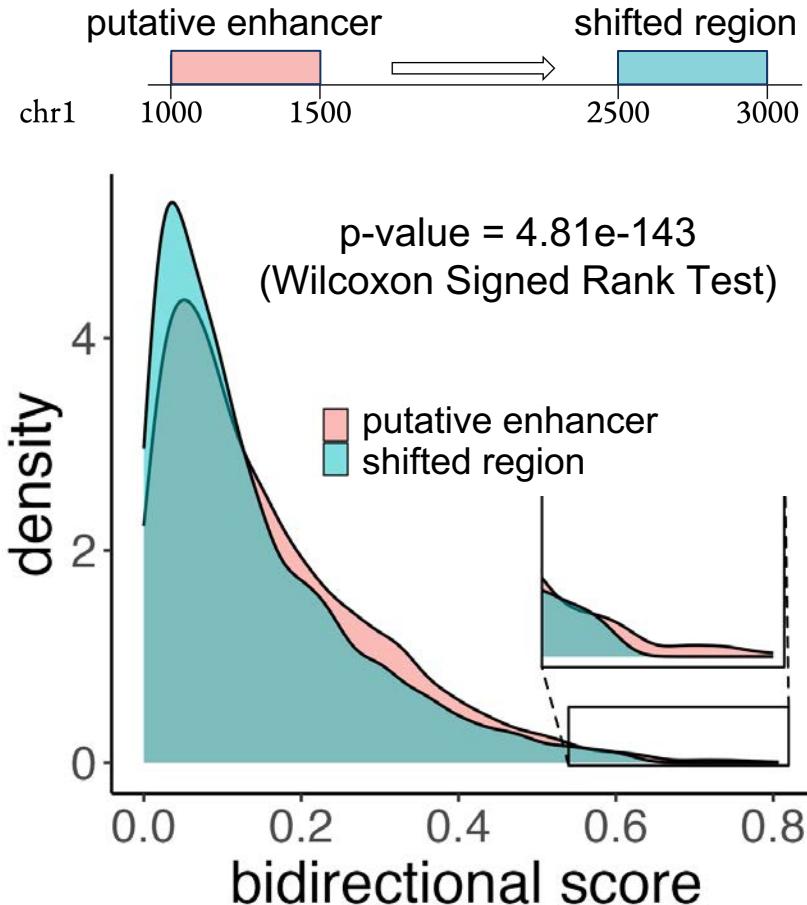
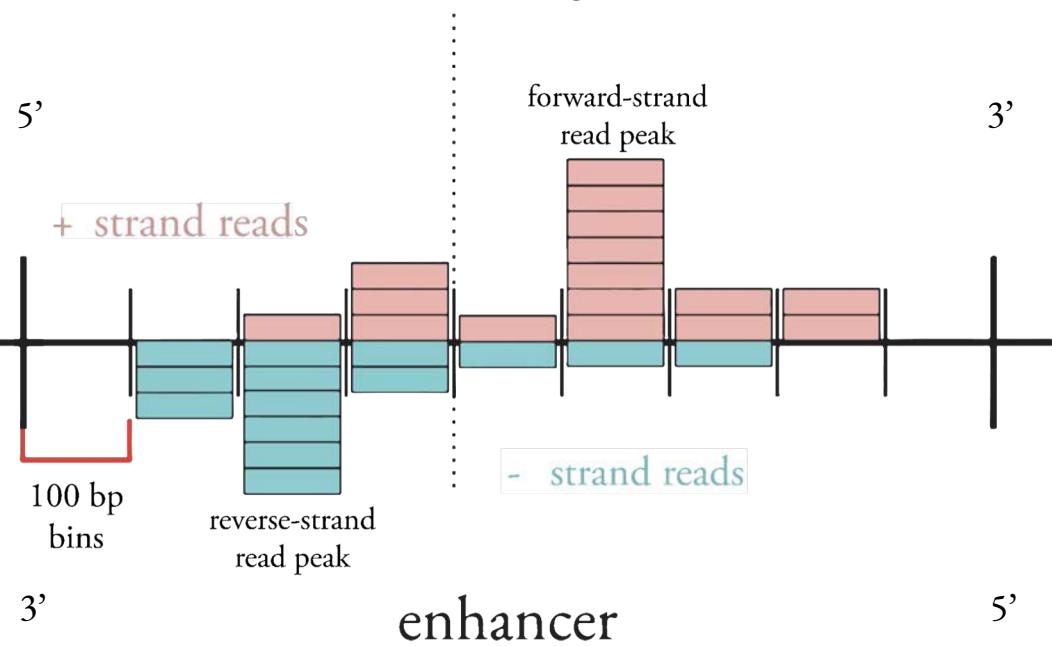


filtered to exclude enhancers that overlapped other regulatory regions

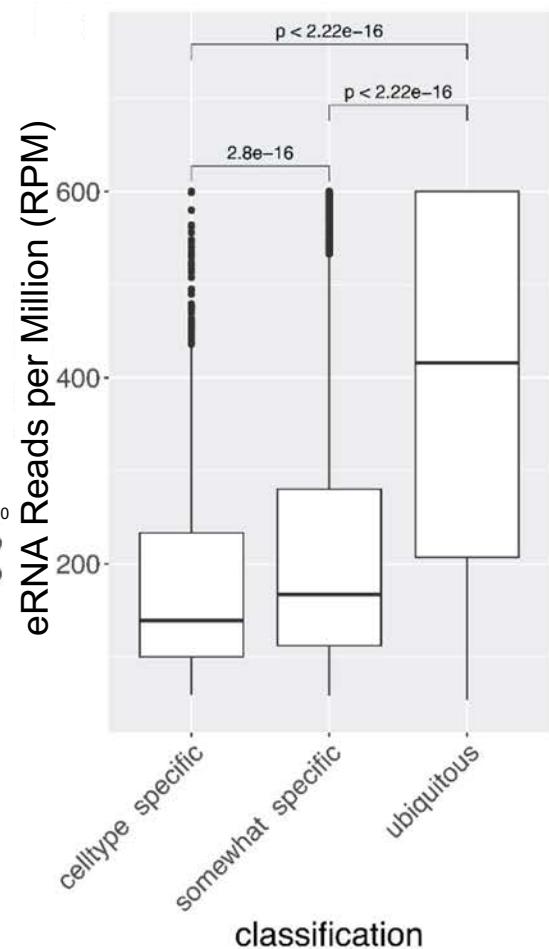
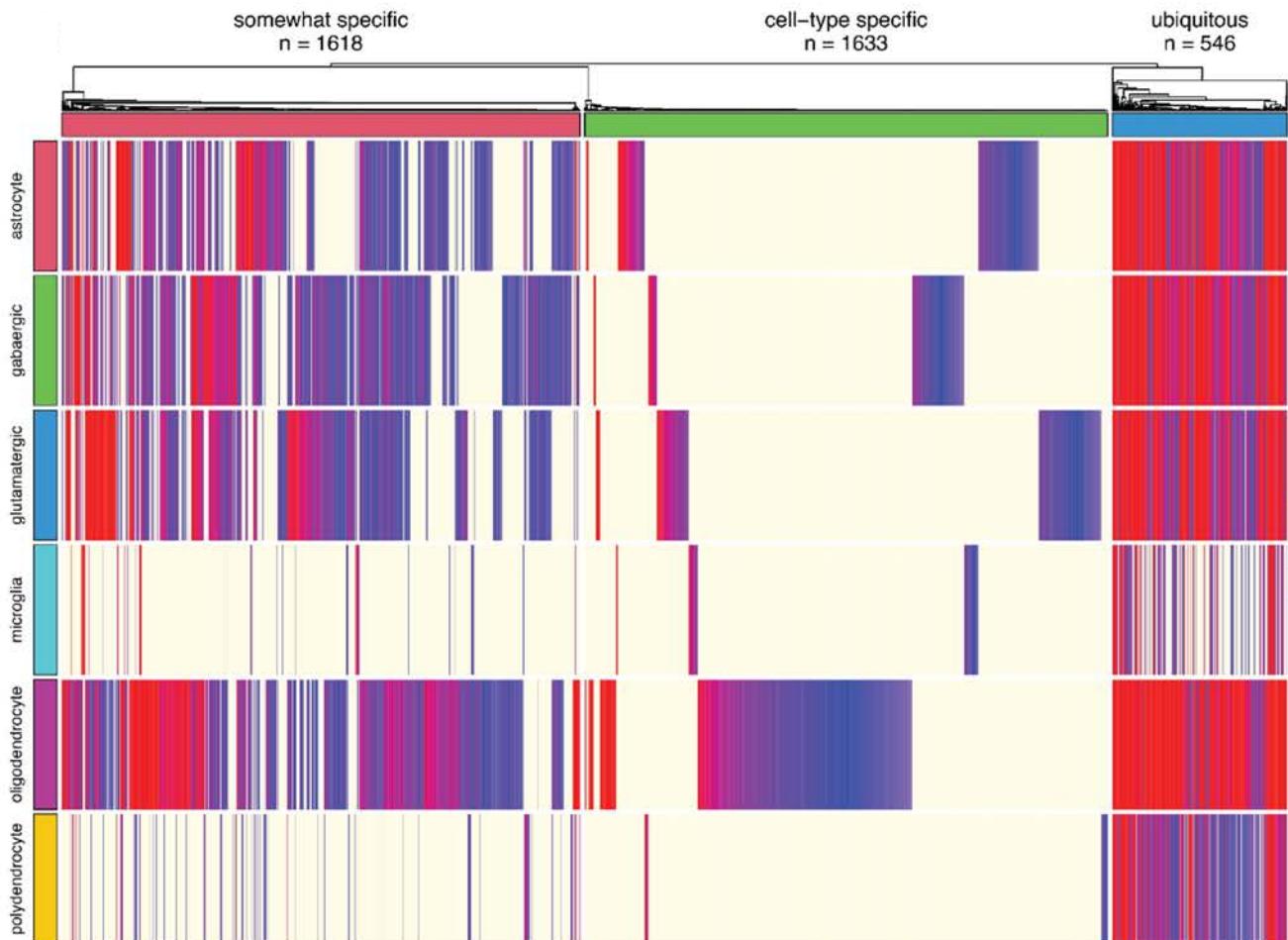
29k putative brain enhancers

Greater bidirectional activity at putative enhancers than nearby intergenic regions

$$S_{bidirectional} = S_{divergence} \cdot (1 - |S_{balanced}|)$$

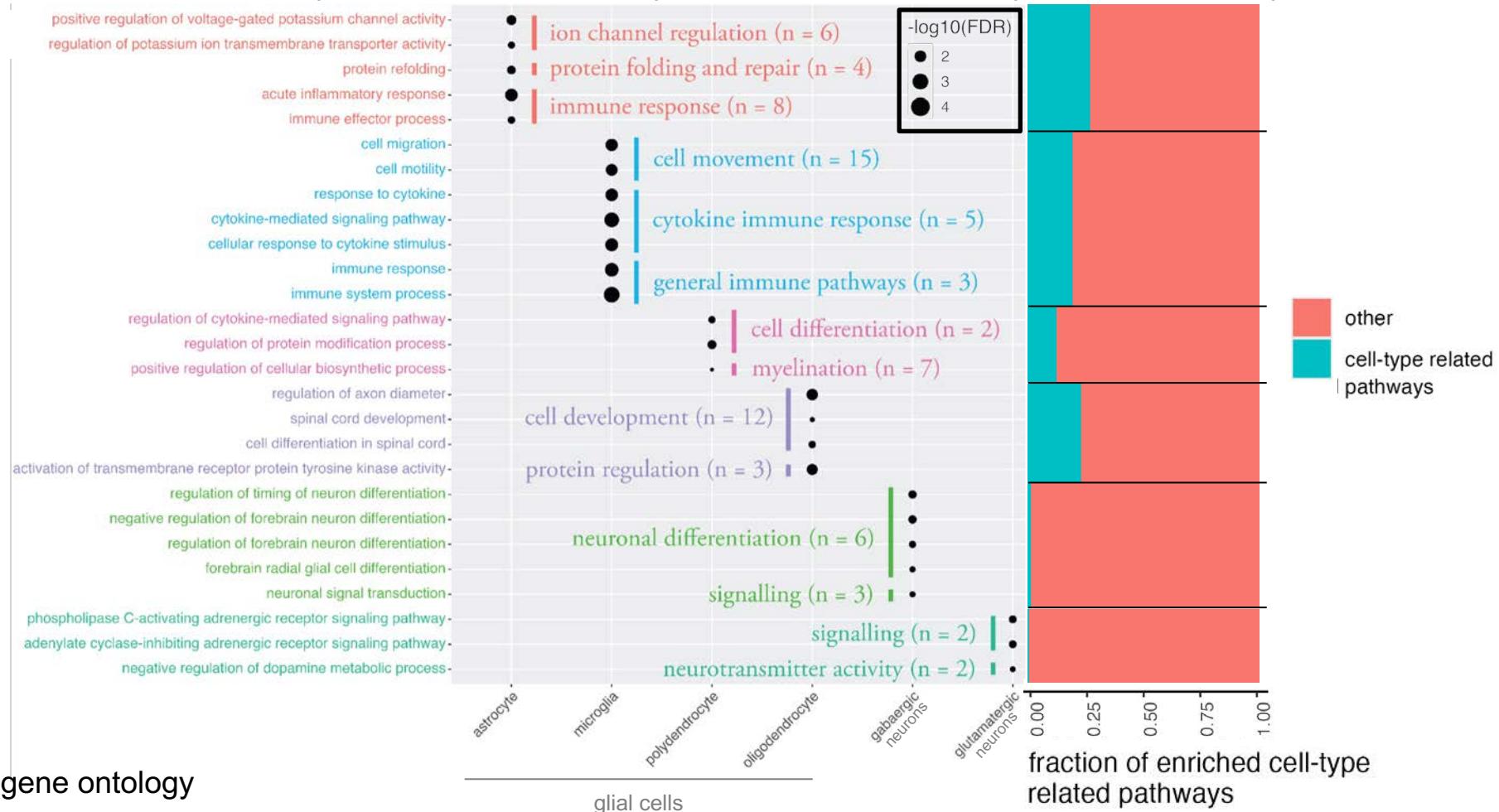


Enhancer expression varies across cell-type



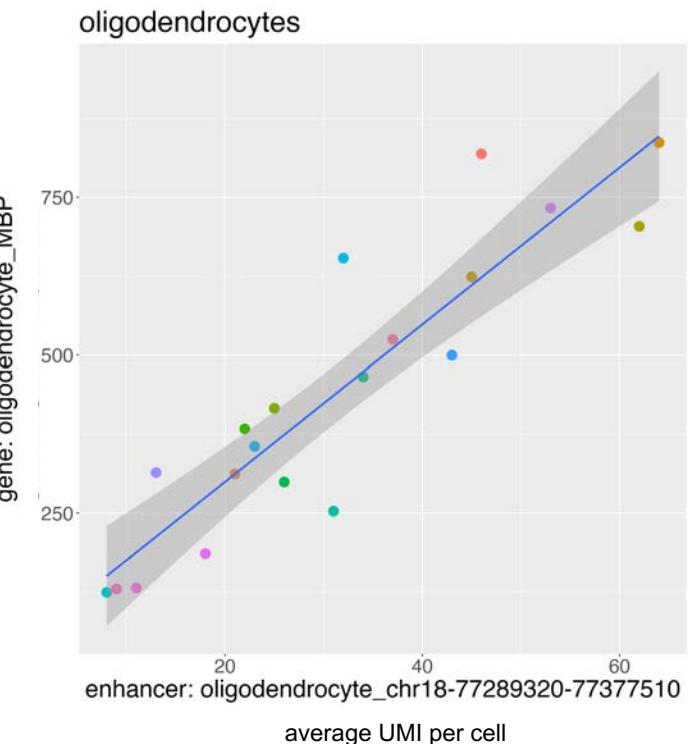
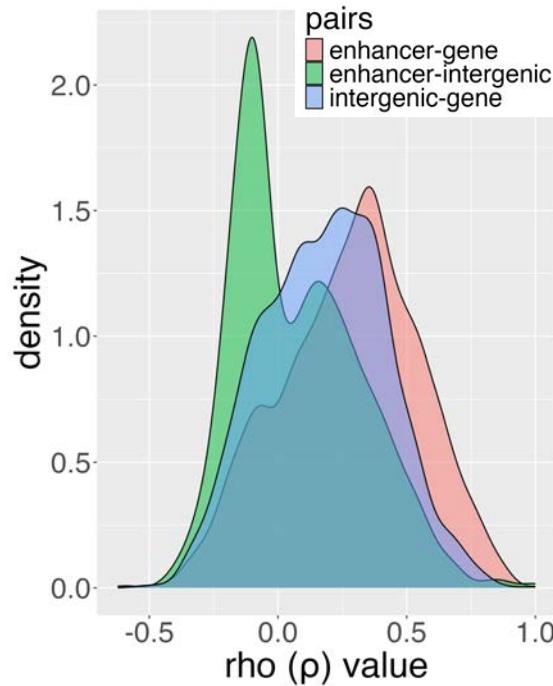
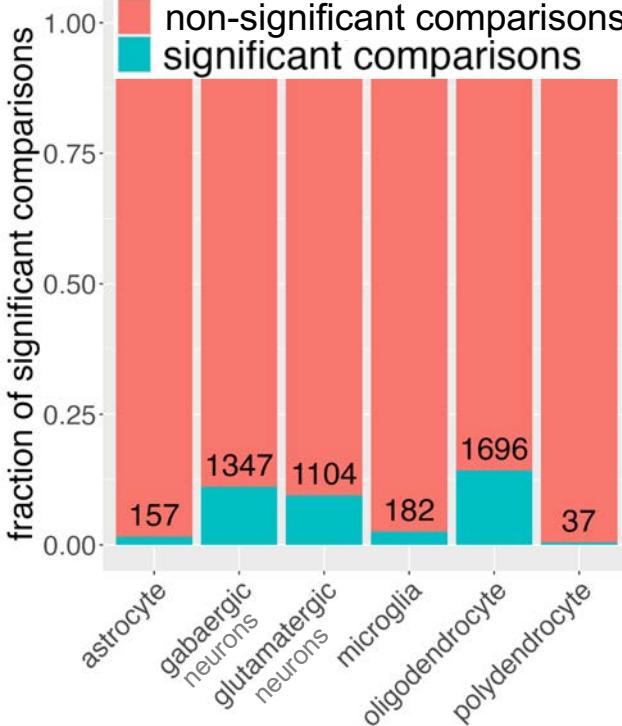
Enhancer cell-type specificity is revealed by GO analysis

Go biological processes



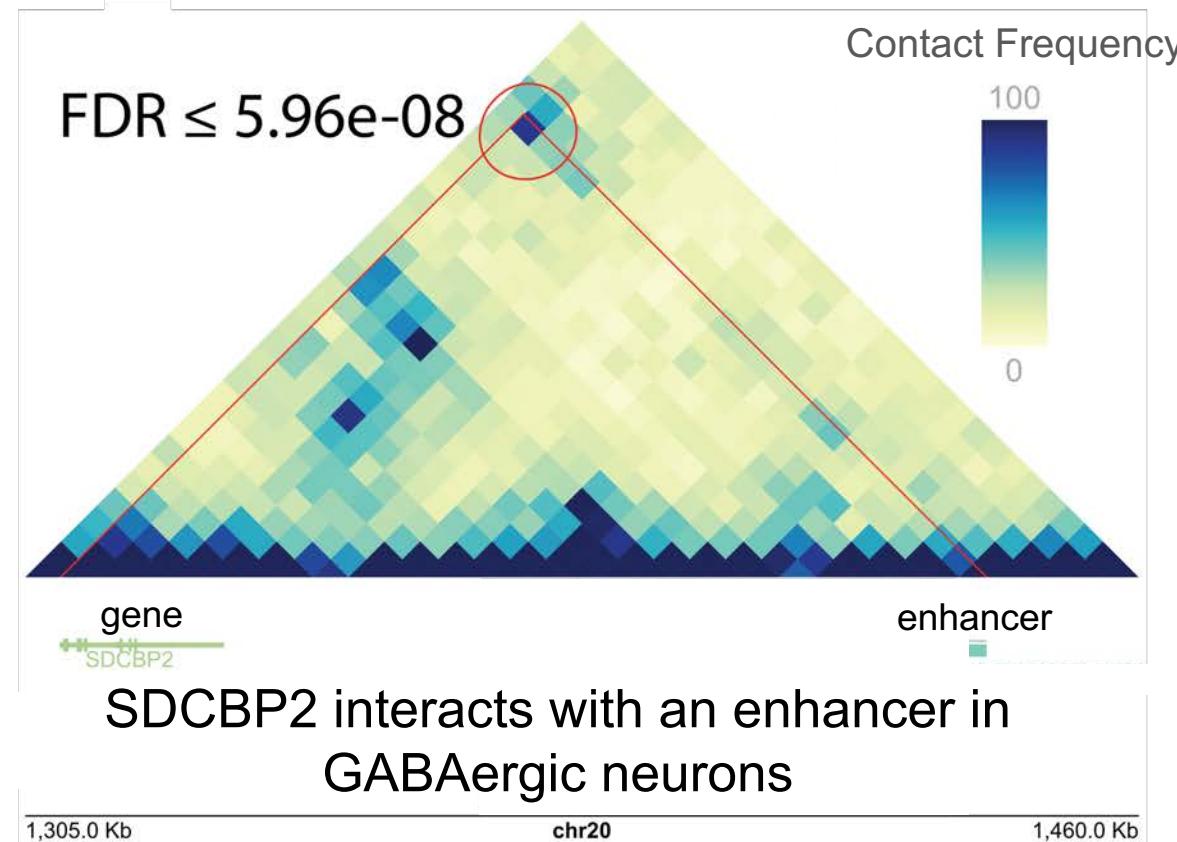
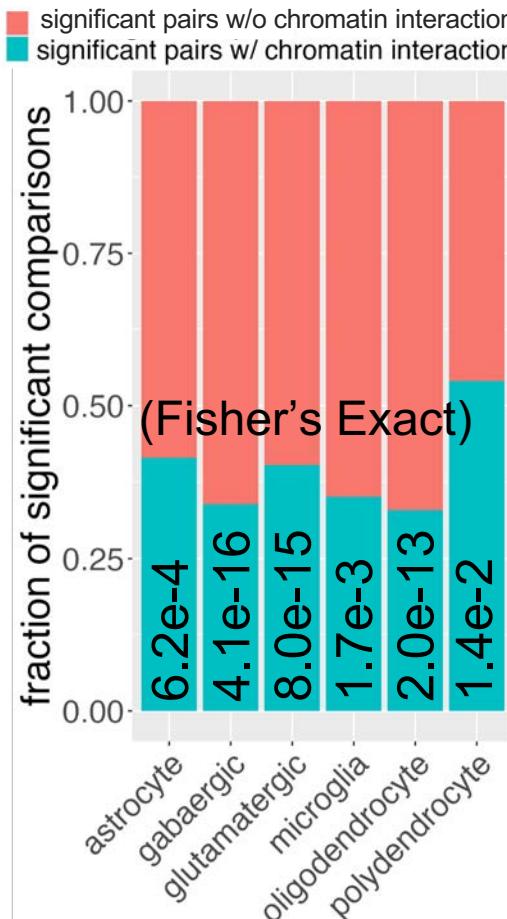
Mapping enhancers to genes

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Enhancer-gene pairs bolstered with Hi-C data

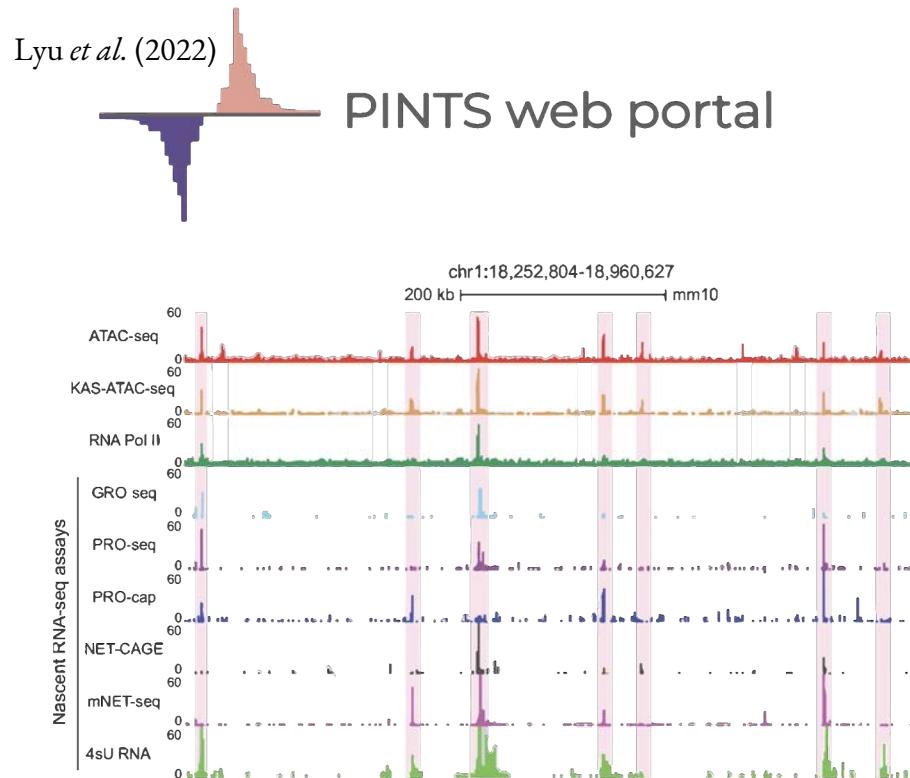
11



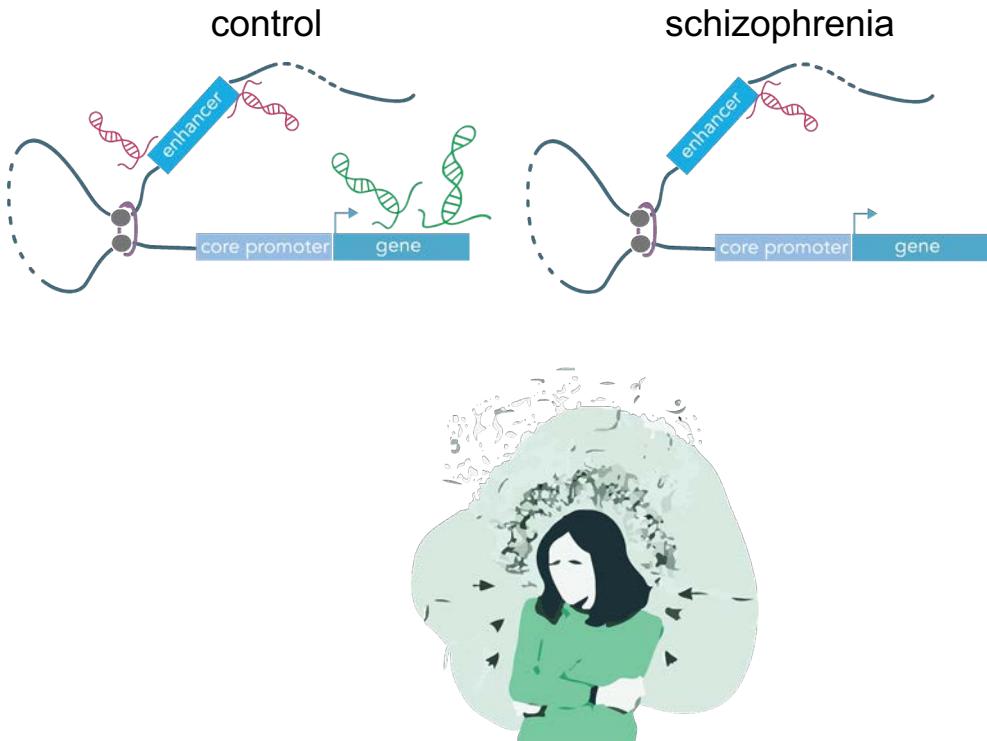
Implications and future plans

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1. Assessing our framework in comparison to other datasets



2. Evaluating the role of our enhancer-gene pairs in diseases



Yao *et al.* (2024)

Artistic image modified from MedKart

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References

1. Theuns, J. et al. Promoter Mutations That Increase Amyloid Precursor-Protein Expression Are Associated with Alzheimer Disease. *Am. J. Hum. Genet.* 78, 936 (2006).
2. Kim, T.-K. et al. Widespread transcription at neuronal activity-regulated enhancers. *Nature* 465, 182–187 (2010).
3. De Santa, F. et al. A Large Fraction of Exogenous RNA Pol II Transcription Sites Overlap Enhancers. *PLoS Biol.* 8, e1000384 (2010).
4. Ren, C. et al. Functional annotation of structural ncRNAs within enhancer RNAs in the human genome: implications for human disease. *Sci. Rep.* 7, 1–15 (2017).
5. Cichewicz, M. A. et al. MUNC, an Enhancer RNA Upstream from the MYOD Gene, Induces a Subgroup of Myogenic Transcripts in trans Independently of MyoD. *Mol. Cell. Biol.* (2018) doi:10.1128/MCB.00655-17.
6. Lam, M. T. Y. et al. Rev-Erbα repress macrophage gene expression by inhibiting enhancer-directed transcription. *Nature* 498, 511–515 (2013).
7. Li, W. et al. Functional roles of enhancer RNAs for oestrogen-dependent transcriptional activation. *Nature* 498, 516–520 (2013).
8. Kaikkonen, M. U. et al. Remodeling of the enhancer landscape during macrophage activation is coupled to enhancer transcription. *Mol. Cell* 51, 310 (2013).
9. eRNAs Are Required for p53-Dependent Enhancer Activity and Gene Transcription. *Mol. Cell* 49, 524–535 (2013).
10. Andersson, R. et al. An atlas of active enhancers across human cell types and tissues. *Nature* 507, 455–461 (2014).
11. Rahnamoun, H., Orozco, P. & Lauberth, S. M. The role of enhancer RNAs in epigenetic regulation of gene expression. *Transcription* 11, 19 (2020).
12. Kristjánsdóttir, K., Dziubek, A., Kang, H. M. & Kwak, H. Population-scale study of eRNA transcription reveals bipartite functional enhancer architecture. *Nat. Commun.* 11, 1–12 (2020).
13. Macosko, E. Z. et al. Highly parallel genome-wide expression profiling of individual cells using nanoliter droplets. *Cell* 161, 1202 (2015).
14. Zhang, Z. et al. Transcriptional landscape and clinical utility of enhancer RNAs for eRNA-targeted therapy in cancer. *Nat. Commun.* 10, 1–12 (2019).
15. Aiden, E. ENCSR165UJN. The ENCODE Data Coordination Center (2022).
16. Lettice, L. A. et al. Disruption of a long-range cis-acting regulator for Shh causes preaxial polydactyly. *Proceedings of the National Academy of Sciences* 99, 7548–7553 (2002).
17. Furlong, E. M. & Levine, M. Developmental enhancers and chromosome topology. *Science* (2018) doi:10.1126/science.aau0320.
18. Mora, A., Sandve, G. K., Gabrielsen, O. S. & Eskeland, R. In the loop: promoter–enhancer interactions and bioinformatics. *Brief. Bioinform.* 17, 980 (2016).
19. Durand, N. C. et al. Juicer Provides a One-Click System for Analyzing Loop-Resolution Hi-C Experiments. *cels* 3, 95–98 (2016).
20. Lyu, R. et al. Quantitative analysis of cis-regulatory elements in transcription with KAS-ATAC-seq. *Nat Commun.* 15, 6852 (2024).
21. Yao, L. et al. A comparison of experimental assays and analytical methods for genome-wide identification of active enhancers. *Nat Biotechnol* 40, 1056–1065 (2022).
22. McCarroll, S., Ling, E. & Goldman, M. Village nuclei isolation with myelin removal v1. (2023) doi:10.17504/protocols.io.4r3l22e3xl1y/v1.
23. Wells, M. F. et al. Natural variation in gene expression and viral susceptibility revealed by neural progenitor cell villages. *Cell Stem Cell* 30, (2023).
24. Ling, E. et al. A concerted neuron–astrocyte program declines in ageing and schizophrenia. *Nature* 627, 604–611 (2024).
25. Alquicira-Hernandez, J., Sathe, A., Ji, H. P., Nguyen, Q. & Powell, J. E. scPred: accurate supervised method for cell type classification from single-cell RNA-seq data. *Genome Biol.* 20, 1–17 (2019).
26. Gao, T. & Qian, J. EnhancerAtlas 2.0: an updated resource with enhancer annotation in 586 tissue/cell types across nine species. *Nucleic Acids Res.* 48, D58–D64 (2019).
27. Amemiya, H. M., Kundaje, A. & Boyle, A. P. The ENCODE Blacklist: Identification of Problematic Regions of the Genome. *Sci. Rep.* 9, 1–5 (2019).
28. Quinlan, A. R. & Hall, I. M. BEDTools: a flexible suite of utilities for comparing genomic features. *Bioinformatics* 26, 841–842 (2010).
29. Frankish, A. et al. GENCODE 2021. *Nucleic Acids Res.* 49, (2021).
30. Ge, S. X., Jung, D. & Yao, R. ShinyGO: a graphical gene-set enrichment tool for animals and plants. *Bioinformatics* 36, 2628 (2020)